



## Original Research Article

# Endoparasitic Helminths of various Species of Frogs in Penang island, Peninsular Malaysia

Wahab A Rahman\* and ZatilShakinah

School of Food Science and Technology Universiti Terengganu Malaysia  
Kuala Terengganu, Malaysia  
\*Corresponding author

## A B S T R A C T

### Keywords

Frogs, Penang Island, Peninsular Malaysia, helminthic parasites

From this study a total of 300 frogs comprising of 14 species were collected from eight study sites in Penang Island, Peninsular Malaysia: *Duttaphrynus melanostictus*, *Phrynomidaspera*, *Hylaranaerythraea*, *H. labialis*, *H. nigrovittata*, *Polypedates leucomystax*, *Fejervary acancrivora*, *F.limnocharis*, *Limnonectesblythii*, *L. paramacrodon*, *L. ibanorum*, *L. ingeri*, *Microhylabutleri*, and *Kaloulapulchra*. Eleven (11) species of helminthes were recovered from the frogs: *Heterakissp.*, *H. vesicularis*, *Trichostrongylus* spp., *Pharyngodon* spp., *Ascaris* spp., *Oswaldocruzia* spp., *Rhabdias* spp., *Glythelminsstaffordi*, *Diplodiscussacculosus*, *Manodistomum* spp., and *Macracanthorynchus* spp.

## Introduction

Taxonomists have described more than 5,000 species of frogs (Duellman and Trueb, 1994). Disease has been a factor in the decline of amphibian populations worldwide, although other factors including habitat loss and fragmentation, chemical pollution, climate changes, introduction of exotic species, increased ultraviolet radiation, and natural pollution have also been responsible for the decline (Hayes *et al.*, 2010). Malaysia is a country with an equatorial climate with high annual humidity ranging from 60% to 90% and rainfalls of 2000 to 3000 mm, resulting with a rich diverse biodiversity of wildlife (Department of Meteorology Malaysia, 2011). Malaysia harbors about 165 species

from six families of anurans of which more than 150 species are found in Borneo (Inger and Stuebing, 2005), 107 species in Peninsular Malaysia, including 26 species in Penang Island (Ibrahim *et al.*, 2008).

As reported by Duellman and Trueb (1986), there are two main threats to amphibians resulting from habitat destruction and environmental pollution. In Malaysia rapid developments for the last fifteen years have led to changes of breeding habitats for survival of amphibians (Ibrahim *et al* 1997). Negative outcomes from forest logging, draining swamps, covering streams of land developments, damming of rivers and draining of irrigation, introduction of weeds and livestock are the main causes that witnessed the inevitable decrease of

amphibian habitats such as swamps, natural waterways, wetlands, forests, rural areas and agricultural lands; secondly is parasitic infection, which has been a concern for the causal decline in frog populations in many parts of the world (Tayler *et al.*, 2007).

In Malaysia there had been some reports of helminthic parasites recovered from frogs. Some nematodes had been described from the intestines of frogs (Fusco and Palmieri 1979; Moravec *et al.*, 2007; Rahman *et al.*, 2008). Other investigators had described several species of trematodes (Yuen, 1961; Rahman *et al.* 2008) and one species of cestode (Ibrahim *et al.*, 1997), also from the intestine of several species of frogs. However, most of these studies were conducted elsewhere in the country, and current knowledge on the helminthic parasites of frogs from the Penang Island, Peninsular Malaysia is lacking.

## **Materials and Methods**

### **(i) Study Sites**

Penang Island, Peninsular Malaysia is located near the northwestern coast of Peninsular Malaysia (5° 24.0' N, 100° 14.0' E). Samplings were carried out during May 2010 to May 2011 from eight (8) areas, randomly selected from various parts of the island.

### **(ii) Sample Collections**

Collection of frogs was carried out at night between 2000h -2300h using torch lights and head lamps. Besides using butterfly nets, frogs were also caught by hand, especially for those frogs found inside rocks crevices. Captured frogs were kept alive in aquarium boxes and transported back to the laboratory.

In the laboratory, the frogs were placed in the sink with running tap water to avoid dehydration of the frog's skin. Species of frogs were identified according to the descriptions of Inger and Stuebing (2005).

Subsequently, frogs were killed by using low doses of chloroform. Frogs were then weighed using a top pan digital electronic balance (SHIMADZU Brand Japan). Frogs were then measured for their snout-vent (cm) and total lengths (cm) by using a ruler scale.

### **Parasites**

Each frog was examined for external abnormalities before the animal was euthanized. The stomach, intestines, rectum, heart and liver were recovered and examined for endoparasites. Blood samples were drawn from the heart.

The frogs as shown in were laid on the dissecting tray with belly facing upwards and their legs were pinned with pins. The first incision was done through the top layer of their skin by using a blade scalpel without damaging the organ. The incision was made vertically down the belly.

Next, the incision was done laterally across the hind legs and across the throat as well. Then the two lateral incisions were connected together with a vertical incision down the belly.

The flaps were opened and pinned them down. Subsequently, the same incision was done on the last layer of frogs' skin by twisting the scissor to avoid cutting organ under the chest bones. The flaps were opened and all fats were removed from the organs.

## Results and Discussion

### Species composition of frogs

Fourteen species of frogs from the five families were identified: *Duttaphrynus melanostictus* and *Phrynomidaspera* (Family Bufonidae); *Fejervarya cancrivora*, *F. limnocharis*, *Limnonectes blythii*, *L. paramacrodon*, *L. ibanorum* and *L. ingeri* (Family Dicroglossidae); *Microhylabutleri* and *Kaloulapulchra* (Family Microhylidae); *Hylarana erythraea*, *H. labialis* and *H. nigrovittata* (Family Ranidae); and *Polypedates leucomystax* (Family Rhacophoridae) (Table 1).

*Duttaphrynus melanostictus* from Bufonidae family seemed to be the most dominant frog species in Penang Island, followed by *H. erythraea* from the Ranidae family. The least common species of frog was *L. ibanorum* from the Dicroglossidae family.

### Composition of helminthes

A total of 8058 individual helminths were collected comprising of 43.7% nematodes, 36.8% trematodes and 19.5% acanthocephalans (Table 2). The most common nematode species found were *Heterakis* spp., *Heterakis vesicularis*, *Pharyngodon* spp., *Trichostrongylus* spp., *Ascaris* spp., *Oswaldocruzias* spp. and *Rhabdias* spp. The three species of trematodes were *Glypthelminis steffordi*, *Diplodiscus sacculousus* and *Manodistomum* spp. One species of acanthocephalan was collected and identified as *Macracanthorhynchus* spp. The frog species *Duttaphrynus melanostictus* seemed to be infected by almost all species of parasites except for the trematode *Manodistomum* spp. Unlike other parasite species, four in particular are confined to three frog host species:

the nematodes *Trichostrongylus* spp. and *Rhabdias* spp. and the trematodes *Diplodiscus sacculousus* and *Manodistomum* spp. The acanthocephalan, *Macracanthorhynchus* spp. seemed to be able to infect numerous species of frog hosts.

### The mean significance between frogs and parasites

One-way ANOVA analysis and LSD test on significant mean differences between the groups of helminthes parasites and frogs for *H. vesicularis*, showed there was high mean significance ( $p < 0.05$ ) in *D. melanostictus* ( $63.14 \pm 24.67$ ), *P. aspera* ( $48.57 \pm 20.20$ ) and *H. erythraea* ( $44.00 \pm 17.06$ ) compared to the other frog species: *P. leucomystax*, *F. cancrivora*, *F. limnocharis*, *M. butleri*, *L. blythii*, *L. ibanorum*, *H. labialis*, *L. paramacrodon*, *L. ingeri*, *K. pulchra* and *H. nigrovittata*. For *Heterakis* spp., there was high significance ( $p < 0.05$ ) in *P. aspera* ( $33.14 \pm 27.03$ ) and *H. erythraea* ( $33.43 \pm 13.31$ ). *Ascaris* spp. was the least number of parasites found in frogs. *P. aspera* ( $29.71 \pm 8.59$ ) had a high significance at level ( $p < 0.05$ ). *Macracanthorhynchus* spp. in the frog *D. melanostictus* ( $120.42 \pm 93.51$ ) had a high mean significance (0.05) followed by *Phrynomidaspera* ( $34.14 \pm 18.51$ ). Conversely, *D. sacculousus* numbers was also found to be low in frogs. But it has a high significance ( $p < 0.05$ ) in *D. melanostictus* ( $16.42 \pm 5.93$ ). *Glypthelminis steffordi* had high significance ( $p < 0.05$ ) in *D. melanostictus* ( $259.85 \pm 213.66$ ), followed by *H. erythraea* ( $78.43 \pm 42.14$ ).

There were 300 frogs from 14 species captured in eight localities in Penang Island, Peninsular Malaysia. Different species of frogs inhabit different habitat areas. Therefore, the distribution of frogs found in every sampling site was different.

**Table.1** Number of frogs caught in eight study sites in Penang Island, Peninsular Malaysia

<b>Frog species</b>	<b>Total numberof frogs caught</b>
<b>Family Bufonidae</b>	
<i>Duttaphrynusmelanostictus</i>	90(27.0)*
<i>Phrynodidaspera</i>	47 (14.1)
<b>Family Ranidae</b>	
<i>Hylaranaerythraea</i>	71(21.3)
<i>Hylaranaraniceps</i>	20 (6.0)
<i>Hylarananigrovittata</i>	2 (0.6)
<b>Family Rhacophoridae</b>	
<i>Polypedatesleucomystax</i>	16 (4.8)
<b>Family Dicroglossidae</b>	
<i>Fejervaryacancrivora</i>	10 (3.0)
<i>Fejervaryalimnocharis</i>	25 (7.5)
<i>Limnonectesblythii</i>	25 (7.5)
<i>Limnonectesparamacrodon</i>	3 (0.9)
<i>Limnonectesibanorum</i>	1 (0.3)
<i>Limnonectesingeri</i>	4 (1.2)
<b>Family Microhylidae</b>	
<i>Microhylabutleri</i>	4 (1.2)
<i>Kaloulapulchra</i>	5 (1.5)
<b>Total</b>	<b>333 (100)</b>

\*Figure in bracket denotes percentage.

**Table.2** Composition of helminthes recovered from the respective frog hosts

Species of helminthshelminthes (%)	Total no. of	Host species
<b><u>Nematode</u></b>		
<i>Heterakis</i> spp.	762 (9.4)	a,b,c,d,e,f, g,i,j,l,n*
<i>Heterakisvesicularis</i>	1345 (16.7)	a,b,c,d,e,f,g, h,i,j,k,l,m,n
<i>Pharyngodon</i> spp.	1021 (12.7)	a,b,c,d,f,g, h,j,k,l,m,n
<i>Trichostrongylus</i> spp.	77 (1.0)	a,c,f
<i>Ascaris</i> spp.	272 (3.3)	a,b,c,d,f,h,i
<i>Oswaldocruzias</i> spp.	31 (0.4)	a,b,d,f,h,j
<i>Rhabdias</i> spp.	13 (0.2)	a,b,n
<b>Percentage</b>	43.7 %	
<b><u>Trematode</u></b>		
<i>Diplodiscussacculousus</i>	144 (1.8)	a,c,j
<i>Glythelminsstaffordi</i>	2601 (32.3)	a,b,c,d,f,j,k,l,m
<i>Manodistomum</i> spp.	214 (2.7)	e,f,j
<b>Percentage</b>	36.8 %	
<b><u>Acanthocephalan</u></b>		
<i>Macracanthorynchus</i> spp.	1578 (19.5)	a,b,c,d,e,f,j,k,m
<b>Percentage</b>	9.5 %	
<b>Total number of helminthes</b>	8058 (100%)	

\*Note: Denotes the respective frog species  
a: *Duttaphrynusmelanostictus*; b:*Phrynoidisaspera*; c: *Hylaranaerythraea*;  
d:*Polypedatesleucomystax*; e:*Fejervaryacancrivora*; f: *F. limnocharis*;  
g: *Microhylabutleri*;h:*Limnonectesblythii*; i: *L. ibanorum*; j: *H. labialis*;  
k: *L. paramacrodon*; l: *Lingeri*; m: *Kaloulapulchra*, n: *H.nigrovittata ingeri*;m:*Kaloulapulchra*; n: *H. nigrovittata*.

*Oswaldocruziaspp.* has not been previously recorded to occur in frog in Malaysia. A previous study discovered *Oxysomatium spp.* and *Rhabdias spp.* from *D. melanostictus* and *F.limnocharis* (Rahman *et al.*, 2008); *Batrachonema synaptospicula* and *Paracosmocera spp* had also been described in *Ranamacrodon spp.* (currently *L. paramacrodon*) (Yuen, 1961; Fusco and Palmieri, 1979) and *Paracapillaria malayensis spp.* from *D.melanostictus* (Moravec *et al.*, 2007).

In a previous study, frog trematodes had been recovered from *Haematoloechu ssingaporensis* from the lungs of *Ranacancrivora* (currently *F.cancrivora*), *D. sacculosus* from the rectums of *R.erythraea* (currently *H. erythraea*) and *G. staffordi* from the intestines of *R. cancrivora*, *R. erythraea* and *R. macrodon*.

There was only one species of acanthocephalan, ie. *Macracanthorhynchus spp.* that was isolated. This species was abundantly found in the intestines of various species of frogs. Most of the parasites were at the larvae stage. A previous study stated that this species was found in pigs and thus the medium of infection mode is unclear. The most common adult form in amphibians is *Acanthocephalus ranae* (Duellman and Trueb, 1986) as well as *A.bufo* (Goldberg *et al.*, 2007). However, so far, not much has been reported on *Macracanthorhynchus spp.* and knowledge of this species is still lacking.

There was no cestode recorded from this study although the study by Ibrahim *et al.*, (1997), reported a species of cestode, *Nematotaenia spp.* in *F. cancrivora* and *F. limnocharis* frogs in northern Peninsular Malaysia.

*Phrynodisaspera* and *H.erythraea* were mostly exposed to water and the chance to get infected by helminthes is higher comparing to other species. As reported by Brooks *et al.* (2006), parasites require water for transmission when the frogs feed on aquatic intermediate hosts or when the parasite is swimming from one host to another host. Besides that, type of habitat also contributes to the parasite infection. A previous study by Kusri *et al.* (2003) reported that the higher prevalence in nematode infection is mostly in terrestrial frogs compared to dwelling frogs. Since *D. melanostictus* is a terrestrial frog, thus explaining that *D.melanostictus* was infected with *H. vesicularis*.

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